

REPORT DOCUMENTATION PAGE			Form Approved OMB NO. 0704-0188		
<p>The public reporting burden for this collection of information is estimated to average 1 hour per response, including the time for reviewing instructions, searching existing data sources, gathering and maintaining the data needed, and completing and reviewing the collection of information. Send comments regarding this burden estimate or any other aspect of this collection of information, including suggestions for reducing this burden, to Washington Headquarters Services, Directorate for Information Operations and Reports, 1215 Jefferson Davis Highway, Suite 1204, Arlington VA, 22202-4302. Respondents should be aware that notwithstanding any other provision of law, no person shall be subject to any penalty for failing to comply with a collection of information if it does not display a currently valid OMB control number.</p> <p>PLEASE DO NOT RETURN YOUR FORM TO THE ABOVE ADDRESS.</p>					
1. REPORT DATE (DD-MM-YYYY)		2. REPORT TYPE New Reprint		3. DATES COVERED (From - To) -	
4. TITLE AND SUBTITLE Thiourea-catalyzed aminolysis of N-acyl homoserine lactones			5a. CONTRACT NUMBER		
			5b. GRANT NUMBER W911NF-04-D-0004		
			5c. PROGRAM ELEMENT NUMBER 611102		
6. AUTHORS Michael A. Bertucci, Stephen J. Lee, Michel R. Gagné			5d. PROJECT NUMBER		
			5e. TASK NUMBER		
			5f. WORK UNIT NUMBER		
7. PERFORMING ORGANIZATION NAMES AND ADDRESSES University of North Carolina - Chapel Hill Office of Sponsored Research 104 Airport Drive, Suite 2200, CB #1350 Chapel Hill, NC 27599 -1350			8. PERFORMING ORGANIZATION REPORT NUMBER		
9. SPONSORING/MONITORING AGENCY NAME(S) AND ADDRESS(ES) U.S. Army Research Office P.O. Box 12211 Research Triangle Park, NC 27709-2211			10. SPONSOR/MONITOR'S ACRONYM(S) ARO		
			11. SPONSOR/MONITOR'S REPORT NUMBER(S) 58118-CH-SR.1		
12. DISTRIBUTION AVAILABILITY STATEMENT Approved for public release; distribution is unlimited.					
13. SUPPLEMENTARY NOTES The views, opinions and/or findings contained in this report are those of the author(s) and should not be construed as an official Department of the Army position, policy or decision, unless so designated by other documentation.					
14. ABSTRACT Thiourea catalysts accelerate aminolysis of N-acyl homoserine lactones (AHLs), molecules integral to bacterial quorum sensing. The catalysts afford rate enhancement of up to 10 times the control in CD3CN. Mild catalysis in other polar aprotic solvents is still observed, while the activity is attenuated in polar protic solvents.					
15. SUBJECT TERMS quorum sensing					
16. SECURITY CLASSIFICATION OF:			17. LIMITATION OF ABSTRACT UU	15. NUMBER OF PAGES	19a. NAME OF RESPONSIBLE PERSON Stephen Lee
a. REPORT UU	b. ABSTRACT UU	c. THIS PAGE UU			19b. TELEPHONE NUMBER 919-549-4365

## **Report Title**

Thiourea-catalyzed aminolysis of N-acyl homoserine lactones

### **ABSTRACT**

Thiourea catalysts accelerate aminolysis of N-acyl homoserine lactones (AHLs), molecules integral to bacterial quorum sensing. The catalysts afford rate enhancement of up to 10 times the control in CD<sub>3</sub>CN. Mild catalysis in other polar aprotic solvents is still observed, while the activity is attenuated in polar protic solvents.

---

**REPORT DOCUMENTATION PAGE (SF298)**  
**(Continuation Sheet)**

---

Continuation for Block 13

ARO Report Number    58118.1-CH-SR  
Thiourea-catalyzed aminolysis of N-acyl homose    ...

Block 13: Supplementary Note

© 2013 . Published in Chemical Communications, Vol. 49 (20) (2013), ( 20). DoD Components reserve a royalty-free, nonexclusive and irrevocable right to reproduce, publish, or otherwise use the work for Federal purposes, and to authorize others to do so (DODGARS §32.36). The views, opinions and/or findings contained in this report are those of the author(s) and should not be construed as an official Department of the Army position, policy or decision, unless so designated by other documentation.

Approved for public release; distribution is unlimited.

Thiourea-catalyzed aminolysis of *N*-acyl homoserine lactones†Cite this: *Chem. Commun.*, 2013, **49**, 2055Received 30th November 2012,  
Accepted 22nd January 2013

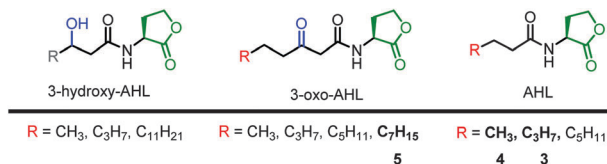
DOI: 10.1039/c3cc00268c

www.rsc.org/chemcomm

**Thiourea catalysts accelerate aminolysis of *N*-acyl homoserine lactones (AHLs), molecules integral to bacterial quorum sensing. The catalysts afford rate enhancement of up to 10 times the control in CD<sub>3</sub>CN. Mild catalysis in other polar aprotic solvents is still observed, while the activity is attenuated in polar protic solvents.**

Translating well-established organic reactions to physiological conditions is one approach for solving contemporary problems in biology. Bacterial virulence mediated by a density-dependent communication pathway known as quorum sensing (QS) is one such problem.<sup>1</sup> For instance, QS pathways in *P. aeruginosa* and *B. cepacia* are responsible for opportunistic infections in immunocompromised patients.<sup>2</sup>

In most gram-negative bacteria, *N*-acyl homoserine lactones (AHLs) are integral to this multicellular communication network (Scheme 1).<sup>3</sup> Efforts to inhibit QS have focused on small molecule inhibitors of the AHL synthases and receptors, while methods of disabling the AHL messenger directly have been less extensively investigated.<sup>4–6</sup> In fact, beyond the physiological degradation of the AHLs, their fundamental reactivity is not well documented.

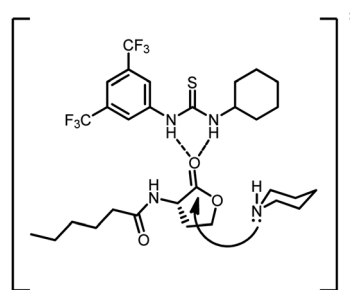


**Scheme 1** AHLs implicated in quorum sensing; sites for interspecies variability include the length of the *N*-acyl chain (red) and the oxidation state (blue). The C<sub>6</sub>-AHL **3**, the C<sub>4</sub>-AHL **4**, and 3-oxo-C<sub>12</sub>-AHL **5** were each tested for amenability to thiourea catalysis.

<sup>a</sup> Department of Chemistry, University of North Carolina at Chapel Hill, Campus Box 3290, Caudill and Kenan Laboratories, Chapel Hill, NC 27599-3290, USA. E-mail: mgagne@unc.edu; Fax: +1 919-962-2388; Tel: +1 919-962-0278

<sup>b</sup> US Army Research Office, P.O. Box 12211, Research Triangle Park, NC 27709, USA

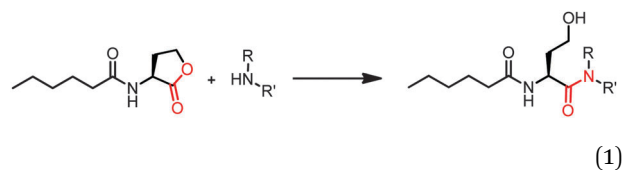
† Electronic supplementary information (ESI) available: Synthetic procedures, <sup>1</sup>H and <sup>13</sup>C NMR spectra, and catalyst screening procedure. See DOI: 10.1039/c3cc00268c

Michael A. Bertucci,<sup>a</sup> Stephen J. Lee<sup>b</sup> and Michel R. Gagné<sup>\*a</sup>

**Scheme 2** The proposed mode of activation of an AHL through H-bonding with a thiourea catalyst.

To begin addressing this deficiency, we sought to determine the susceptibility of AHLs to nucleophilic ring opening, specifically by amines to form stable amides (eqn (1)).

Traditionally, reactions of amines with butyrolactones are sluggish and require a large excess of the nucleophile and/or heat.<sup>7</sup> The recalcitrant nature of this simple acyl transfer reaction offered the opportunity to develop catalysts for the aminolysis of AHLs. Use of thiourea organocatalysts appeared fitting as they have proven effective in the supramolecular activation of esters (Scheme 2).<sup>8</sup> This H-bond activation mechanism has been showcased in a variety of classic organic methodologies, including the polymerization of cyclic esters.<sup>9</sup> Of relevance to potential biological applications are examples of thiourea catalysis in water.<sup>10</sup> In this communication, the utility of thiourea catalysts for the aminolysis of AHLs is assessed to determine if these catalysts can be utilized for irreversible AHL derivitization.



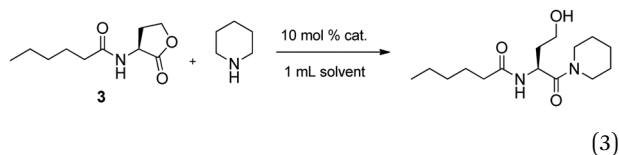
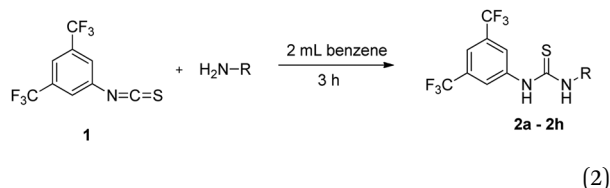
To evaluate the ability of thiourea organocatalysts to accelerate the aminolysis of AHLs, we synthesized a small library of catalysts (Table 1). The thioureas were assembled

**Table 1** Yields for the synthesis of each thiourea catalyst and relative rate constants for 10 mol% of the catalysts in aminolysis of the C<sub>6</sub>-AHL with piperidine<sup>a</sup>

Catalyst	R	Yield (%)	<i>k</i> <sub>rel</sub> CD <sub>3</sub> CN	<i>k</i> <sub>rel</sub> DMF-d <sub>7</sub>
2a		90	4.5	3.4
2b		84	5.6	3.6
2c		82	4.3	3.4
2d		87	4.4	3.3
2e		38	5.5	3.2
2f		11	4.5	—
2g		79	7.9	5.2
2h		83	10	5.3

<sup>a</sup> For further details on the screening procedure, see ESI. The reactions were monitored until 50% conversion (*t*<sub>50</sub>) was reached. See Table 3 for the *t*<sub>50</sub> values of the uncatalyzed reaction.

through the reaction of a primary amine with 3,5-bis(trifluoromethyl) phenylisothiocyanate (**1**) to form **2** (eqn (2)). The electron withdrawing ability of the 3,5-bis(trifluoromethyl) phenyl functionality is known to enhance the H-bond donating ability and consequent catalytic potency of the thiourea.<sup>11</sup> Eight potential organocatalysts were synthesized; monofunctional catalysts **2a** and **2b** from simple alkyl amines, and bifunctional catalysts **2c–2f** from alkylated diamines and histamine. The scope of bifunctional catalysts ranged from catalysts hypothesized to assist in nucleophile activation by H-bonding interactions (**2c** and **2d**) to those conceivably providing anionic transition state stabilization (**2f**).<sup>9,12</sup> The reaction of simple diamines with **1** yielded dithiourea compounds **2g** and **2h**.



The catalysts were then screened for activity in a C<sub>6</sub>-AHL (**3**) (50 mM) and piperidine (20 eq.) control reaction (eqn 3).<sup>†</sup> AHL conversion in each catalytic trial was monitored by <sup>1</sup>H NMR spectroscopy in CD<sub>3</sub>CN. Prior to the addition of piperidine, a slight downfield shift in the N–H resonances of the thiourea

was observed, supporting an AHL–catalyst interaction. The entire library of thioureas proved active in accelerating the aminolysis of the AHL. The monofunctional and bifunctional thioureas enhanced the nucleophilic ring opening up to 5.6 times the background reaction while the dithioureas reached a rate 10-fold faster than the control. Notably, the bifunctional catalysts did not provide any noticeable advantage compared to the monofunctional catalysts. This lack of secondary activation may be due to an inherent weakness of the anticipated N–H···N bonding interaction between the amine nucleophile and the nitrogen containing appendages of the catalysts.

Given the structural variety in the AHL family, two other AHLs were tested. The C<sub>4</sub>-AHL (**4**) and 3-oxo-C<sub>12</sub>-AHL (**5**) are both utilized by the human pathogen *P. aeruginosa*, but vary in alkyl chain length; the 3-oxo-C<sub>12</sub>-AHL is also oxidized at C<sub>3</sub> of the alkyl chain (Scheme 1).<sup>3</sup> Both AHLs were screened in CD<sub>3</sub>CN in the presence of catalyst **2a** and our best performing catalyst **2h** (10 mol%) (Table 2). The C<sub>4</sub>-AHL performed comparably to the C<sub>6</sub>-AHL. On the other hand, the barely soluble 3-oxo-C<sub>12</sub>-AHL was less amenable to thiourea catalysis with **2a** and **2h**, displaying a rate increase only 2.4 and 4.0 times the control, respectively. Though lipophilic interactions could be leading to substrate aggregation in the markedly nonpolar 3-oxo-C<sub>12</sub>-AHL, the decrease in relative rate may also be attributed to the extra carbonyl moiety's competitive affinity for the thiourea.

The effect of solvent on catalysis and thermal background reactivity was also assessed (Table 3). In general, more polar solvents decelerated the aminolysis rate (entries 1–5), though catalytic rate accelerations were still observed. Catalyst **2h** was quite effective in DMF-d<sub>7</sub> (entry 4), a solvent that we expected might deactivate the catalyst by competitive H-bonding.

To progress closer to physiological conditions, a variety of protic co-solvents (10% v/v) were tested. These protic additives

**Table 2** Relative rates of aminolysis of different AHLs in CD<sub>3</sub>CN<sup>a</sup>

AHL	<i>k</i> <sub>rel</sub> ( <b>2a</b> )	<i>k</i> <sub>rel</sub> ( <b>2h</b> )
C <sub>6</sub> -AHL ( <b>3</b> )	4.5	10
C <sub>4</sub> -AHL ( <b>4</b> )	4.1	7.3
3-oxo-C <sub>12</sub> -AHL ( <b>5</b> )	2.7	4.0

<sup>a</sup> With piperidine as the amine nucleophile in 20-fold excess.

**Table 3** Relative rates of C<sub>6</sub>-AHL aminolysis in the presence of catalysts **2a** and **2h** in aprotic solvents and protic solvent mixtures<sup>a</sup>

Entry	Solvent (% v/v)	<i>t</i> <sub>50</sub> <sup>b</sup> uncat (×10 <sup>3</sup> s)	<i>k</i> <sub>rel</sub> ( <b>2a</b> )	<i>k</i> <sub>rel</sub> ( <b>2h</b> )
1	C <sub>6</sub> D <sub>6</sub>	5.1	2.0	4.3
2	CD <sub>2</sub> Cl <sub>2</sub>	3.6	1.0	1.1
3	CD <sub>3</sub> CN	35.5	4.5	10
4	DMF-d <sub>7</sub>	100.7	3.4	5.3
5	DMSO-d <sub>6</sub>	123.2	1.9	2.2
6	10% MeOH-d <sub>4</sub> -CD <sub>3</sub> CN	14.3	0.9	1.5
7	10% D <sub>2</sub> O-DMSO-d <sub>6</sub>	21.0	1.3	1.0
8	10% TFE-d <sub>3</sub> -CD <sub>3</sub> CN	9.7	1.1	1.0
9	10% D <sub>2</sub> O-CD <sub>3</sub> CN	4.2	1.0	0.9

<sup>a</sup> With piperidine as the amine nucleophile in 20-fold excess.

<sup>b</sup> *t*<sub>50</sub> represents the reaction time to reach 50% conversion.

dramatically accelerated the aminolysis reactions, but negated the effects of the added catalysts (entries 6–9, Table 3). Overall, increasing solvent polarity served to decrease background reaction rates, while protic additives increased them. It seems reasonable to suggest that in addition to accelerating the background reaction, the protic nature of the co-solvents simultaneously deactivate the catalyst by inhibiting the necessary supramolecular events that have been proposed for thiourea catalysis (Scheme 2).<sup>9–12</sup>

Taking simple, well-establish synthetics methodologies and applying them to biologically relevant targets helps assess the utility of those methods in a physiological setting. In the present case, thioureas have been demonstrated to provide up to 10-fold rate accelerations for the aminolysis of AHLs. However, attempts to transition these observations to conditions that are more physiological (polar protic) reveal the significant challenge of using small molecule catalysts in such environments. Nevertheless, information of AHL fundamental reactivity has emerged from these studies.

We wish to acknowledge Dr Masa Matsumoto, Dr Silvia Bezer-Bratu and Dr Mee-Kyung Chung for their valuable insights and offer a special thanks to Dr Marcey L. Waters and the US Army Research Office for their support.

## Notes and references

† Piperidine was chosen during a screen of several amines for reaction with C<sub>6</sub>-AHL. The timescale of the reaction was appropriate for monitoring aminolysis by <sup>1</sup>H NMR.

- 1 General background on bacterial quorum sensing: C. M. Waters and B. L. Bassler, *Annu. Rev. Cell Dev. Biol.*, 2005, **21**, 319–346; S. R. Chhabra, B. Philipp, L. Eberl, M. Givskov, P. Williams and M. Cámara, *Top. Curr. Chem.*, 2005, **240**, 279–315; S. Uroz, Y. Dessaux and P. Oger, *ChemBioChem*, 2009, **10**, 205–216; S. T. Rutherford and B. L. Bassler, *Cold Spring Harbor Perspect. Med.*, 2012, 1–25.
- 2 M. Cámara, P. Williams and A. Hardman, *Lancet Infect. Dis.*, 2002, **2**, 667–676; L. C. M. Antunes, R. B. R. Ferreira, M. M. C. Buckner and B. B. Finlay, *Microbiology*, 2010, **156**, 2271–2282.
- 3 N-acyl homoserine lactones: S. R. Chhabra, C. Harty, D. S. W. Hooi, M. Daykin, P. Williams, G. Telford, D. J. Pritchard and B. W. Bycroft, *J. Med. Chem.*, 2002, **46**, 97–104; A. M. Pomini and A. J. Marsaioli, *J. Nat. Prod.*, 2008, **71**, 1032–1036; M. E. A. Churchill and L. Chen, *Chem. Rev.*, 2011, **111**, 68–85.
- 4 W. R. J. D. Galloway, J. T. Hodgkinson, S. D. Bowden, M. Welch and D. R. Spring, *Chem. Rev.*, 2011, **111**, 28–67.
- 5 Small molecule quorum sensing inhibitors: K. M. Smith, Y. Bu and H. Suga, *Chem. Biol.*, 2003, **10**, 563–571; F. G. Glansdorp, G. L. Thomas, J. K. Lee, J. M. Dutton, G. P. C. Salmond, M. Welch and D. R. Spring, *Org. Biomol. Chem.*, 2004, **2**, 3329–3336; G. D. Geske, R. J. Wezeman, A. P. Siegel and H. E. Blackwell, *J. Am. Chem. Soc.*, 2005, **127**, 12762–12763; T. B. Rasmussen and M. Givskov, *Microbiology*, 2006, **152**, 895–904; G. D. Geske, J. C. O'Neill, D. M. Miller, M. E. Mattmann and H. E. Blackwell, *J. Am. Chem. Soc.*, 2007, **129**, 13613–13625.
- 6 Enzyme, antibody mediated quorum quenching: Y. Lin, J. Xu, J. Hu, L. Wang, S. L. Ong, J. R. Leadbetter and L. Zhang, *Mol. Microbiol.*, 2003, **47**, 849–860; G. F. Kaufmann, R. Sartorio, S. Lee, J. M. Mee, L. J. Altobelli III, D. P. Kujawa, E. Jefferies, B. Clapham, M. M. Meijler and K. D. Janda, *J. Am. Chem. Soc.*, 2006, **128**, 2802–2803; S. De Lamo Marin, Y. Xu, M. M. Meijler and K. D. Janda, *Bioorg. Med. Chem. Lett.*, 2007, **17**, 1549–1552; J. F. Teiber, S. Horke, D. C. Haines, P. K. Chowdhary, J. Xiao, G. L. Kramer, R. W. Haley and D. I. Draganov, *Infect. Immun.*, 2008, **76**, 2512–2519; P. B. Kapadnis, E. Hall, M. Ramstedt, W. R. J. D. Galloway, M. Welch and D. R. Spring, *Chem. Commun.*, 2009, 538–540; N. Amara, B. P. Krom, G. F. Kaufmann and M. M. Meijler, *Chem. Rev.*, 2011, **111**, 195–208.
- 7 G. Blay, L. Cardona, B. Garcia, C. L. Garcia and J. R. Pedro, *Tetrahedron Lett.*, 1994, **35**, 931–934; W. Liu, D. D. Xu, O. Repič and T. L. Blacklock, *Tetrahedron Lett.*, 2001, **42**, 2439–2441.
- 8 P. R. Schreiner, *Chem. Soc. Rev.*, 2003, **32**, 289–296; J. Connon, *Chem.-Eur. J.*, 2006, **12**, 5418–5427.
- 9 A. P. Dove, R. C. Pratt, B. G. G. Lohmeijer, R. M. Waymouth and J. L. Hedrick, *J. Am. Chem. Soc.*, 2005, **127**, 13798–13799.
- 10 A. Wittkopp and P. R. Schreiner, *Chem.-Eur. J.*, 2003, **9**, 407–414; C. M. Kleiner and P. R. Schreiner, *Chem. Commun.*, 2006, 4315; Y. Cao, Y. Lai, X. Wang, Y. Li and W. Xiao, *Chem. Inform.*, 2007, **38**, 15.
- 11 K. M. Lippert, K. Hof, D. Gerbig, D. Ley, H. Hausmann, S. Guenther and P. R. Schreiner, *Eur. J. Org. Chem.*, 2012, 5919–5927.
- 12 Y. Takemoto, *Chem. Pharm. Bull.*, 2010, **58**, 593–601.